

CORRELATION BETWEEN *YERSINIA* SPP. FOUND IN FAECES AND *BRUCELLA SUIIS* INFECTION IN WILD BOARS *

**Carla Grattarola ¹, Maria Silvia Gennero ¹, Stefania Bergagna ¹,
Simona Zoppi ¹, Laura Chiavacci ² et Alessandro Dondo ¹**

SUMMARY: 379 wild boar faeces samples were collected from July 2005 to June 2006 and tested to detect *Yersinia* spp.. The same animals were submitted to bacteriological and serological tests to detect *Brucella* infection.

62 *Yersinia* spp. strains were isolated. Out of them, 31 were identified as *Y. enterocolitica*.

The χ^2 test has showed neither correlation between *Yersinia* spp. and serological tests ($\chi^2 = 0.009$, $p=0.97$) nor between *Yersinia* spp. and *Brucella suis* isolation ($\chi^2 = 0.03$, $p=0.87$).

Contrary to what occurs in cattle, seropositivity in naturally infected wild boars may reflect infection by *Brucella suis* rather than by *Yersinia* spp.

Keywords: *Yersinia* spp., crossreactivity, *Brucella suis*.

RESUME : La recherche de *Yersinia* spp a été faite dans 379 fèces de sanglier récoltées entre Juillet 2005 et Juin 2006 ; chaque animal a été soumis à des tests de bactériologie et de sérologie pour confirmer l'infection par *Brucella*. Soixante deux souches de *Yersinia* spp. ont été isolées et 31 ont été identifiées comme *Y. enterocolitica*.

Le χ^2 test n'a démontré aucune corrélation entre l'isolement de *Y. enterocolitica* et les tests sérologiques ($\chi^2 = 0,009$, $p=0,97$) ou entre l'isolement de *Yersinia* spp. et celui de *Brucella suis*.

Au contraire de ce qu'on observe chez les bovins, on peut supposer que chez le sanglier la seropositivité brucellique est liée à l'infection par *Brucella suis* et non pas à celle par *Yersinia* spp.

Mots-clés: *Yersinia* spp., réaction croisée, *Brucella suis*.



I - INTRODUCTION

Brucellosis and yersiniosis are diseases which affect animals as well as human beings.

Brucella infection caused by *Brucella suis* is a zoonosis common to different animal species and although *B. suis* (biovar 1, 2 and 3) is still

widely distributed in the world, prevalence in domestic pigs is low, with the exception of South-East Asia and South America [Godfroid J., 2002].

* Texte de la communication orale présentée lors de la Journée AEEMA, 1^{er} juin 2007

¹ Istituto Zooprofilattico Sperimentale di Piemonte, Liguria e Valle d'Aosta, Via Bologna 148, 10154 Torino, Italy; carla.grattarola@izsto.it

² Osservatorio Epidemiologico Regione Piemonte

In Central Europe, including Italy, this infection is endemic; brown hare (*Lepus europaeus*) and wild boar (*Sus scrofa*) are the natural reservoirs of *B. suis* biovar 2.

Yersiniosis has been described in many species of domestic and free-living mammals and birds. Recently, several outbreaks of this emerging food-borne disease have been reported.

In wild boar infectious agents are *Yersinia pseudotuberculosis* and *Yersinia enterocolitica* and particularly, *Y. enterocolitica* O:9 shows similarity to the O-chain of *Brucella* spp., producing false seropositivity versus *Brucella* spp., as shown in particular in cattle [Garin-Bastuji B. *et al.*, 1999; Godfroid J. *et al.*, 2002].

The purpose of this work was to study the correlation between *Yersinia* spp. detection in faeces and serological response to *Brucella* spp. in naturally infected wild boar.

II - MATERIALS AND METHODS

1. SAMPLING

The complete test protocol includes sampling sera, organs (spleen and/or genital organs) and faeces from 379 wild boar hunted from July 1, 2005 to June 30, 2006 in areas where *Brucella suis* infection is endemic.

2. SEROLOGY

We used a technique described by Alton [Alton *et al.*, 1988] and optimised by the Italian National Institute of Health based on the Rose Bengal Test (RB) and the Complement Fixation Test (CFT).

3. BRUCELLA SUIIS ISOLATION FROM ORGANS

Bacteriological isolation was also performed on animal tissues (uterus, spleen and testicles). 20 g of tissue were macerated in 25 ml of sterile saline in a Stomacher® (International Pbi). The whole material was inoculated into modified Thayer-Martin medium [Farrell, 1974; Martin *et al.*, 1996] and *Brucella* medium developed by Farrell [AA.VV., 2004], enriched with 10% horse serum and selective supplement and incubated at 37°C in an atmosphere containing 5% CO₂ for at least 10 days. All colonies resembling *Brucella* were seeded into blood agar medium and incubated

for a further 2 days before re-examination. If *Brucella* was suspected with Stamp's staining [Alton *et al.*, 1988], the colonies were then identified to species by classical techniques [Alton *et al.*, 1967].

Isolated strains were seeded and typed by biochemical [Alton *et al.*, 1988] and molecular tests [Bricker *et al.*, 1994; Cloeckaert *et al.*, 1995; Corbel *et al.*, 1983].

4. ISOLA-TION OF Y. ENTEROCOLITICA FROM FAECES

About 5 g of faeces were sent to the laboratory and tested. Phosphate buffered saline (PBS) was added to the samples in a ratio of 1:20 v/v.

The culture broths were incubated at 4°C for 21 days and weekly seeded on selective agar (CIN). In order to evaluate lactose fermentation and to proceed with the biochemical studies, if a suspected colony developed the colony was transplanted to MacConkey agar and blood agar and the plates were incubated at 30°C for 24 h. Urease study was performed on suspected colonies; in the presence of positive urease, identification was performed with a miniaturized API 20 E gallery. The strains confirmed as *Yersinia enterocolitica* were serotyped with monovalent antisera for somatic antigens.

III - RESULTS

A total of 62 faecal samples out of 379 gave positive results for *Yersinia* spp. 31 out of them were identified as *Y. enterocolitica* and then serotyped. The remaining samples were positive to *Y. kristensenii* (26), *Y. frederiksenii* (2), *Y. aldovae* (2) and *Y. intermedia* (1).

Serotyping has led to identify 12 strains of *Y. enterocolitica* (YE) O:9, 5 YE O: 5, 4 YE O:8 and 1 YE O:1,2; 9 strains didn't give any conclusive result.

The table below (table 1) shows the results of the serological and bacteriological tests. RB and CFT have been evaluated in parallel.

For purposes of statistical analysis we considered only cases in which *Yersinia* spp. were isolated from faecal samples (*Yersinia* positive/negative). Subsequently in order to evaluate the presence of *Yersinia enterocolitica* in wild boars, all strains were identified by means of API gallery [Stockes, 2000].

Table 1
Results and relative comparison of serological and bacteriological tests
(N: negative; P: positive)

<i>Brucella</i> spp.		Yersinia negative	YE +	Yersinia + (other than YE)	Total	%
Serological test	Bacteriological test					
N(299)	N(292)	244	25	23	48	16.4% (48/292)
	P(7)	6	1	0	1	14.2% (1/7)
P(80)	N(67)	56	4	7	11	16.4% (11/67)
	P(13)	11	1	1	2	15.3% (2/13)
Total	379	317	31	31	62	

IV - DISCUSSION

In wild boar groups, examined in different areas where *Brucella* infection is endemic, we should assume that seropositivity is most likely linked to the presence of *Brucella* spp. Table 2 shows *Yersinia* spp. detection has been indifferently obtained both from seropositive (16.2%) and seronegative animals (16.7%).

Chi-square test shows that *Yersinia* spp. detection in faeces and seropositivity to *Brucella* spp. are independent ($\chi^2 = 0.009$, $p=0.97$).

Similarly, we have observed no correlation between *Yersinia* spp. and *Brucella suis* isolation ($\chi^2 = 0.03$, $p=0.87$).

Thus, the agreement between *Yersinia* detection and serological/bacteriological test is poor, which strengthens the hypothesis that *Yersinia* presence is independent from the two considered tests, differently than observed in cattle.

Table 2
Results and relative comparison of serological test and *Yersinia* spp. isolation
(N: negative; P: positive)

<i>Brucella</i> spp. serological response	<i>Yersinia</i> spp. isolation from / examined samples
N(299)	16.7% (50/299)
P(80)	16.2% (13/80)

BIBLIOGRAPHY

- AA VV - Manual of Standards for Diagnostic Test and Vaccines. OIE, 2004.
- Alton G.G., Jones M.L. - Laboratory techniques in *Brucella*. WHO, 1967.
- Alton G.G., Jones M.L., Angus R.D., Verger J.M. - Techniques for the brucellosis laboratory. INRA, Paris, 1988.
- Bricker B., Halling S.M. - Differentiation of *Brucella abortus* bv. 1,2 and 4, *Brucella melitensis*, *Brucella ovis* and *Brucella suis* bv. 1 by PCR. *JCM.*, 1994, **32**, 2660-2666.
- Cloekaert A., Verger J.M., Grayon M., Grepinet O. - Restriction site polymorphism of the genes encoding the major 25 kDa and 36 kDa outer membrane proteins of *Brucella*. *Microbiol.*, 1995, **141**, 2111-2121.
- Corbel M.J., Thomas E.L. - The *Brucella* phages their properties characterisation and application Research Section Disease of Breeding Department-central Veterinary Laboratory, Weybridge, 1983.
- Dahouk S., Nockler K., Tomaso H., Splettstoesser W.D., Jungersen G., Riber U., Petry T., Hoffmann D., Scholz H.C., Hensel A., Neubauer H. - Seroprevalence of Brucellosis, Tularemia, and Yersiniosis in Wild Boars (*Sus scrofa*) from North-Eastern Germany. *J. Vet. Med. B.*, 2005, **52**, 444-455.
- Davis D.S. - Animal Brucellosis Boca Raton, FL : CRC Press Inc., 1990, 321-324.
- Dondo A., Grattarola C., Gennero M.S., Zoppi S., Di Giannatale E. - Osservazioni preliminari sulla presenza di *Brucella suis* biovar 1 nel cinghiale in Piemonte. *Il Progresso Veterinario*, 2003, **3**, 112-116.
- Farrell I.D. - The development of new selective medium for the isolation of *Brucella abortus* from contaminated sources. *Res. Vet. Sci.*, 1994, **16**, 280-286.
- Garin-Bastuji B. - Brucellose du sanglier. *Bull. Inf. Path. Anim. Sauv.*, 1997, **17**, 17-18.
- Garin-Bastuji B., Hummel H., Gerbier G., Cau C., Pouillot R., Da Costa M., Fontaine J-J. - Non specific serological reactions in the diagnosis of bovine brucellosis: experimental oral infection of cattle with repeated doses of *Yersinia enterocolitica* O:9. *Vet. Microbiol.*, 1994, **66**, 223-233.
- Godfroid J., Michel P., Uytterhaegen L., De Smedt C., Rasseneur F., Boelaert F., Saegerman C., Patigny X. - Brucellose enzootique à *Brucella suis* biotype 2 chez le sanglier (*Sus scrofa*) en Belgique. *Ann. Med. Vet.*, 1994, **138**, 263-268.
- Godfroid J. - Brucellosis in wildlife. *Rev. sci. tech. Off. Epiz.*, 2002, **21** (2), 277-286.
- Godfroid J., Saegerman C., Wellemans V., Walravens K., Letesson J-J., Tibor A., Millan A.M.C., Spencer S., Sanna M., Bakker D., Pouillot R., Garin-Bastuji B. - How to substantiate eradication of bovine brucellosis when aspecific serological reactions occur in the course of brucellosis testing. *Vet. Microbiol.*, 2002, **90**, 461-477.
- Hars J., Valery M., Chaduc F., Garin-Bastuji B., Pinguet O., Rossi S. - Surveillance de la Brucellose du sanglier et du lièvre dans le département de l'Allier. *BIPAS*, 2000, **23**, 121-138.

Martin C.M., Alabart J.L., Blasco J.M. - Effect
of antibiotics contained in two Brucella
selective media on growth of *B.abortus*, *B.*

melitensis and *B.ovis*. *J. Clin. Microbiol.*,
1996, **34**, 426-428.

Stockes M.E. *et al.* - Categorical Data Analysis
Using The SAS System 2nd Edition, 2000.

